



Application

Comparative evaluation of FastGene® Scriptase Basic and other company's kit by RT-PCR

Category

Reverse Transcriptase

Product

FastGene® Scriptase Basic (LS52)

Manufacturer

NIPPON Genetics EUROPE

The following data was published due to the kindness of Mr. Tamura, a member of the Japanese public research institute.

Overview



FastGene® Scriptase Basic (LS52)

In order to evaluate the performance of FastGene® Scriptase Basic, reverse transcription was performed on the RNA extracted from HeLA cells with this product and other company's reverse transcriptase.

The obtained cDNA was amplified by PCR for the target gene and confirmed by electrophoresis. The performance of FastGene® Scriptase Basic was evaluated by comparing the band signals.

Result

RNA extraction

Sample: HeLa cells were cultured on a 35 mm dish and used at 70-80% confluency.

RNA extraction: Ribozol (MS Technosystems) (N580-100ML)

Absorbance measurement: U-3900 (HITACHI) (U-3900)

Result: A260/280 = 1.56, yield = $9.5 \mu g$

• Reverse transcription reaction

Total RNA was diluted to 1, 10, 100 ng/ μ L. 1 μ L of each dilution was added as Template RNA. $\,\%$ 0 ng is 1 μ L H $_2$ O

Primer

Gene specific primer (GAPDH): CTCTTCCTCTTGTGCTCTTGC

Template RNA (0, 1, 10, 100 ng/µL) : 1 µL Gene specific primer (GAPDH) : 2 μL (2 pmol) 42 °C 5 min add 10X FastGene® Scriptase buffer : 2 µL dNTP Mixture (2 mM each) : 2 µL FastGene® Scriptase basic : 1 µL Sterile water (RNase free) : 12 µL total : 20 µL 1 42 °C 60 min 90 °C 5 min keep at 22 ℃

Reference: Template total RNA recommended amount: 1 ng-5 µg

company T, RT rection kit

```
Template RNA (0, 1, 10, 100 ng/µL)
                                         : 1 µL
Gene specific primer (GAPDH)
                                         : 2 μL (2 pmol)
2 mM dNTP
                                         : 5 µL
RNase free dH<sub>2</sub>O
                                         : 2 µL
                                         : 10 µL
total
65 °C 5 min
4 °C 5 min
add
                                         : 4 µL
5X buffer
Reverse transcriptase
                                         : 1 µL
RNase free dH<sub>2</sub>O
                                         : 5 µL
42 °C 50 min
70 °C for 15 min
keep at 22 ℃
```

Reference: Template total RNA recommended amount: 5 µg or less



PCR conditions

Reaction composition			
10 μM Forward Primer	:	2.5	μL
10 μM Reverse Primer	:	2.5	μL
2 mM dNTPs (TOYOBO)	:	5	μL
5X Phusion HF Buffer	:	10	μL
cDNA	:	2	μL
sterile water	:	27.5	μL
Phusion DNA Polymerase (NEB)	:	0.5	μL
total	:	50	ul

• Prog	ram		
95 ℃	2 min		
\downarrow			
95 ℃	10 sec	\neg	
65 °C	30 sec		30 cycles
72 ℃	30 sec	\perp	•
\downarrow			
72 °C	3 min		
1			

hold 22 ℃

• Primers for-GAPDH

Forward Primer: CCACAGTCCATGCCATCAC Reverse Primer: CCATGAGGTCCACCACCC

Amplification product size: 500bp

Electrophoresis condition

Agarose gel : 1% agarose gel

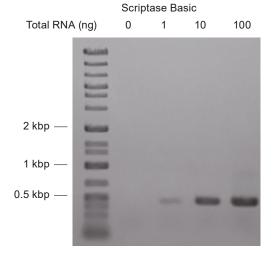
Sample : PCR reaction solution (15 µL)

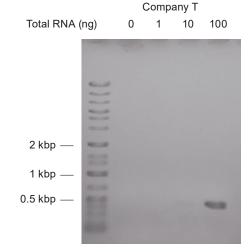
Running buffer : 1xTAE

Migration conditions : 100 V, 30 min

Marker : 7 μL (Gene Ladder Wide 1) (Nippon Gene) (Code No. 313-06961)

Result





% 0 ng is1 $\mu L~H_2O$

We were able to detect PCR bands by using Scriptase Basic and a small amount of total RNA.



Scriptase Basic is relatively inexpensive, so it is cost effective and satisfactory. The tube labels are color-coded, which makes them easy to use.

Customer's comment