



Application

Evaluation of the FastGene[®] miRNA Enhancer kit with the FastGene[®] RNA Premium kit or other companie's RNA extraction kits

\subset	Product
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G	/anufacturer

FastGene[®] miRNA Enhancer kit (Cat.No. FG-RNAE-S, FG-RNAE-25) FastGene[®] RNA Premium kit (Cat.No. FG-81006, FG-81050, FG-81250) FastGene[®]

The following data is kindly provided by Mr. Arizumi Kikuchi. Institute of Medical Science, General Hospital Daiyukai, Japan

Summary

Analyzed samples are from clinical specimens such as tissue, blood, urine and cerebrospinal fluid. It is necessary to remove PCR reaction inhibitors to ensure a sufficient amount of highly purified RNA.

Extraction of nucleic acids from a biological sample in a genetic test is extremely important for maintaining the accuracy of the test. In particular, it is essential to construct an extraction method that enables stable sample preparation for clinical samples that are expected to have various conditions. In this application note, we combined the FastGene® miRNA Enhancer kit with the FastGene® RNA Premium kit or other supplier RNA extraction kit to extract miRNAs from various clinical samples. We evaluated the Cp value by real-time PCR after cDNA synthesis.

• What are miRNAs?

• Approximately 30% of human genes are expected to be regulated by miRNAs.

• After transcription from the genome, the precursor miRNA becomes a mature miRNA after sevaral enzymatic interactions

- mature miRNAs contain 21-23 nt single-stranded RNAs that interact with mRNAs.
- According to miRBase Release22 (2018), 38,589 are registered.



Involvement has been suggested for cancer, metabolic diseases, neurological and infectious diseases



Target sample and pretreatment method

· Cell line (K562)

 \rightarrow Use pellets of 10⁵ cells (n=3)

• human white blood cell (clinical specimen)

 \rightarrow Blood was collected from 3 healthy subjects using a blood collection tube with EDTA \cdot 2Na anticoagulate.

- Use white blood cell pellets after treatment with 500 μL of whole blood (n=3)
- Formalin-fixed paraffin-embedded (FFPE) tissue sample (clinical sample)
- \rightarrow FFPE sample of colorectal tissue collected from 5 patients with colorectal cancer (* for details)
- Bovine muscle
 - \rightarrow 10 μL of homogenized solution using the same amount of PBS (-) as the tissue (n=3)



Meltzer, P.S. Nature 435, 745-746 (2005) .

*Details of FFPE pretreatment method

- 1) Take a sample from a 5 µm thick FFPE specimen
- 2) Add 800 µL xylene
- 3) Incubate for 5 min at room temperature
- 4) Add 400 µL absolute ethanol
 - 5) Centrifuge for 2 min and discard the supernatant
 - 6) Add 1000 µL absolute ethanol
 - 7) Centrifuge for 2 min and discard the supernatant
 - 8) Incubate at 56 $^{\circ}$ for 20 min
 - Add 100 μL dissolution buffer (10 nmol/L Tris-HCl, 0.1 mmol/L EDTA, 5 g/L SDS)
 - 10) Add 40 μ L of proteinase K
 - 11) Incubate at 56 $^\circ\!C$ for 30 min
 - 12) Incubate at 85 $^\circ\!C$ for 30 min
 - 13) Add lysis buffer to make the total volume of 150 μ L







Real-time PCR conditions

Stem-loop RT primer and cDNA synthesis used for miR-21 analysis (used in Cell line • human white blood cell • FFPE sample)

• Stem- Loop RT-primer sequences $5' \rightarrow 3'$				 Reaction condition 			
<u>miR-21</u>	16 ℃ 30 min						
GTCAGAGGAGGTGCAGGGTCCGAGGTATTCGCACCTCCTCTGACTCAACA				30 ℃ 30 s —	٦		
				42 ℃ 30 s 60 cycl	es		
• Reverse Transcription \rightarrow cDNA				50 ℃ 1 s —			
Transcriptor RT Reaction Buffer (5×) (Roche)	2	μL		85 ℃ 5 min			
Stem-Loop RT-primer(10 μM)	0.2	μL					
Transcriptor Reverse Transcriptase (20 U/µL) (Roche) Protector Rnase Inhibitor (40 U/µL) (Roche)		μL		Dilute the reaction product 5 fold with TE buf	upt 5 fold with TE buffor		
		μL		and used for subsequent reactions	ont reactions		
Deoxynucleotide Mix (10 mM) (Roche)	1	μL		and used for subsequent reactions			
RNA solution	2.5	μL					
Water	3.8	μL					

Primer sequence, probe, reaction conditions and reaction solution during real-time PCR used for miR-21 analysis

 Primer sequences (5'→3') and Probe <u>miR-21</u> Forward primer GATCGGTAGCTTATCAGACTGATG Reverse primer GTGCAGGGTCCGGGTAAT Universal ProbeLibrary Probe (Roche) #82

Reaction mixtures
2.5 µL of cDNA solution
5 µL of Essential Probe Master (Roche)
0.4 µM of each primer
0.4 µM of UPL probe (Roche)
(in a final volume of 10 µL)

Reaction conditions
 95 °C 10 min
 95 °C 10 sec _____ 40 cycles
 60 °C 30 sec _____

The reaction was performed with LightCycler 96 (Roche). The average value of the double meassurement was used as the measured value.

Stem-loop RT primer and cDNA synthesis used for bta-miR-23a analysis (used in Bovine muscle)

 Stem- Loop RT-primer sequences 5'→3' 				 Reaction condition 			
bta-miR-23a				16 °C	30 min		
CTCAACTGGTGTCGTGGAGTCGGCAATTCAGTTGAGTGGAAATC *				30 ℃	30 s —	7	
*Guan L et. al.:Sci Rep, 7: 43716, 2017				42 ℃	30 s	60 cycles	
• Reverse Transcript \rightarrow cDNA				50 ℃	1 s —		
Transcriptor RT Reaction Buffer (5×) (Roche)	2	μL		85 ℃	5 min		
Stem-Loop RT-primer (10µM)	0.2	μL					
Transcriptor Reverse Transcriptase (20 U/µL) (Roche) Protector Rnase Inhibitor (40 U/µL) (Roche)		5 μL 5 μL	•	Dilute the reaction product 5-fold with TE buffer, and used for subsequent reactions			
							Deoxynucleotide Mix (10 mM) (Roche)
RNA solution	2.5	μL					
Water	3.8	μL					

Primer sequence, reaction conditions and reaction solution used for bta-miR-23a analysis

• 1 Primer sequences (5'→3') *	 Reaction conditions 		
bta-miR-23a Forward primer_CCGAGTCAGATCACATTGCCAGG	95 ℃	10 min	
Reverse primer CTCAACTGGTGTCGTGGAGTCG	95 ℃ 60 ℃	$\frac{10 \text{ sec}}{30 \text{ sec}} = 40 \text{ cycles}$	
*Guan L et. al. :Sci Rep, 7: 43716, 2017			
Reaction mixtures			
2.5 μL of cDNA solution			
5 μL of KAPA SYBR Fast qPCR(KAPA)			
0.4 µM of each primer			

(in a final volume of 10 μL)

The reaction was performed with LightCycler 96 (Roche). The average value of the double meassurement was used as the measured value.



Result

• Using Protocol I or II : Evaluation of Cp value by real-time PCR while using FastGene® miRNA Enhancer or not





Conclusion: The addition of FastGene® miRNA Enhancer improved miRNA yields in all kits

• Usage of Protocol | and II:

Real-time PCR when using FastGene® miRNA Enhancer by Evaluation of Cp value



Conclusion: The Cp values are the same when using protocol I and II

• Using protocol I and III: Evaluation of ∆Cp



 $\Delta Cp = (Cp \text{ value when FastGene}^{\circ} \text{ miRNA Enhancer is not used}) - (Cp value when FastGene}^{\circ} \text{ miRNA Enhancer is used})$

•Summary The addition of FastGene[®] miRNA Enhancer improved miRNA yields with either kit. It was also found that the amount of extracted miRNA was about the same even if the number of times the column was passed was different.



In our laboratory, we have experience in extracting miRNA using FFPE and peritoneal dialysate drainage as samples, but there were still cases where it was difficult to extract a sufficient amount for analysis. In such a case, I thought that the use of this reagent would be effective.

Since the FastGene[®] series RNA extraction reagents have excellent purity and yield, we believe that combining them with these reagents can be expected to be as effective as or better than the miRNA extraction reagents of other companies.

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 Usage of protocol III: Evaluation of Cp value by real-time PCR when using various samples



Conclusion: The yield of miRNA was improved by using protocol III