

User Guide

4-Channel Standard Quantitative Cartridge Kit (C405101-Q)

A. Specifications

Specification	Description
DNA Sizing Range	20-5,000 bp
DNA Sizing Precision*	2% CV
DNA Quantitative Precision*	4% CV
Sample Number	800 runs**
Shelf Life	6 months

* Best resolution is determined by the 15-622 DNA Size Marker (C109200).
** 200 runs/channel x 4 channels = 800 runs

B. Kit Components and Storage Conditions

Item	Storage Condition
4-Channel Standard Quantitative Cartridge (C405101-Q)	4-30°C (Do Not Freeze)
20-1,500 bp Quantitative Marker (C109109-500Q, 500 µL)	Short-Term (≤ 3 months): 4-30°C Long-Term (> 3 months): -20°C
15-622 bp Size Marker (C109200-100, 100 µL)	Short-Term (≤ 3 months): 4-30°C Long-Term (> 3 months): -20°C
Separation Buffer (C104409, 100 mL)	4-30°C
Dilution Buffer (C104408, 30 mL)	4-30°C
Mineral Oil (C104407, 15 mL)	4-30°C

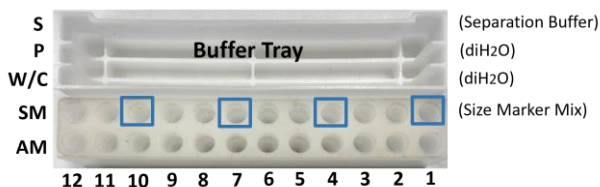
• Please always store cartridges in a light-proof bag, and then store in the cartridge box after analysis.

C. Cartridge Unpacking Preparation

The new cartridge must undergo HV check and calibration. Please place 4 tubes of 20 µL C109109 Quantitative Marker at positions AM01, AM04, AM07, and AM10, and follow the instructions provided in the unpacking guide to complete the calibration process.

- Calibration mix for cartridge unpacking:
 - 20 bp-1,500 bp Quantitative Marker (C109109): 5 µL
 - Dilution Buffer (C104405): 15 µL

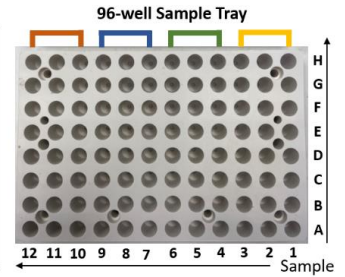
D. Buffer, Marker, and Sample Preparation



4-Channel position:

(1) Channel 1 : A1-H1/A2-H2/A3-H3
 (2) Channel 2 : A4-H4/A5-H5/A6-H6
 (3) Channel 3 : A7-H7/A8-H8/A9-H9
 (4) Channel 4 : A10-H10/A11-H11/A12-H12

*All 4 channels will be analyzed simultaneously.
 *Sample should be in quadruple number. Otherwise, tubes with dilution buffer should be used.



The 4-channel system can be interpreted as 4 individual cartridges. Thus, the sample number should be quadruple.

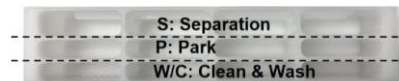
For example, if 4 samples are assigned at A1, A4, A7, and A10, then 6 samples can be assigned at A1, A4, A7, A10, B1, and B4, with dilution buffer at B7 and B10.

1. Compatible Sample Tubes

Name	Cat. No.	Volume	Image
0.1 mL PCR Tube	-	≥ 10 µL	
0.2 mL PCR Tube	-	≥ 20 µL	
Semi/Non-skirted 96-well Plate*	-	10-20 µL	

* Recommended 96-well plates: Labcon (3972-520-000), Axygen (PCR-96-FLT-C).

2. Buffer Preparation



Use the droppers to fill the wells. Each well should be 80% full. Overfilling or having droplets left on the dividers will conduct the currents and make it hard to keep track of the changes.

3. Size Marker Mix Preparation

- For Sample Size Range from 20 bp to 1,500 bp:
 - 20 bp-1,500 bp Quantitative Marker (C109109): 5 µL
 - 15-622 bp Size Marker (C109200): 10 µL
 - Dilution Buffer (C104405): 5 µL
- For Sample Size Range from 20 bp to 5,000 bp:
 - 20 bp-5,000 bp Quantitative Marker (C109102): 5 µL
 - 50 bp-3,000 bp Size Marker (C109301): 10 µL
 - Dilution Buffer (C104405): 5 µL

4. Sample Mix Preparation

- 20 bp-1,500 bp Quantitative Marker (C109109): 5 µL
- Sample: X (2~15) µL
- Dilution Buffer (C104405): 15-X (0~13) µL

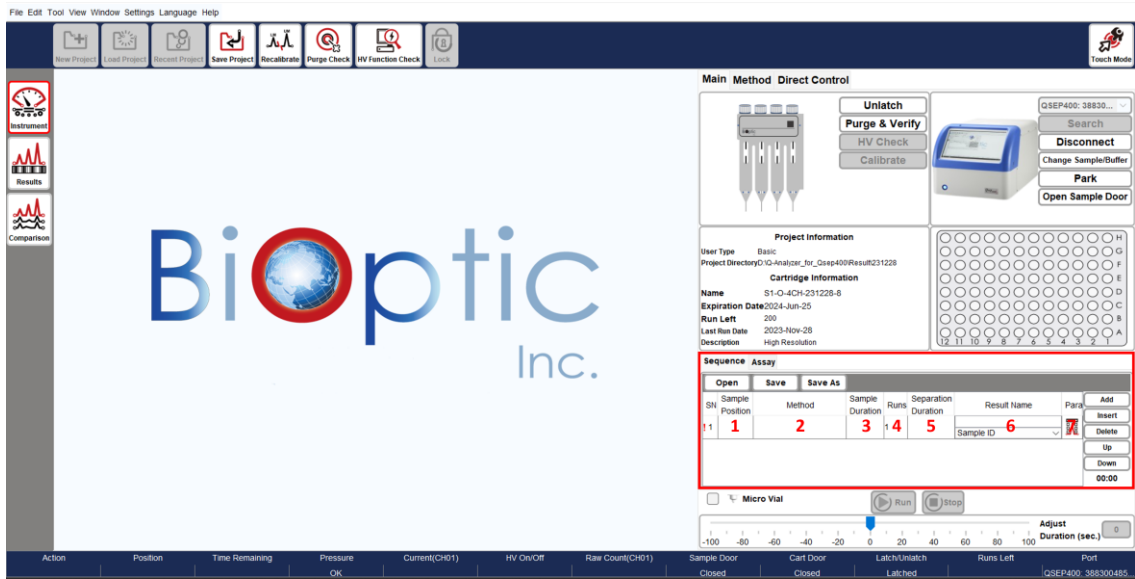
5. Recommended Sample Concentration

Fragmented sample: 0.2-50 ng/µL [Best: 0.5-5 ng/µL]
 Smear sample: 1-50 ng/µL [Best: 2-20 ng/µL]

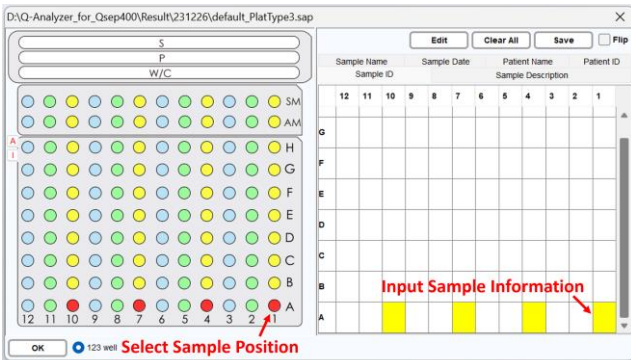
User Guide

4-Channel Standard Quantitative Cartridge Kit (C405101-Q)

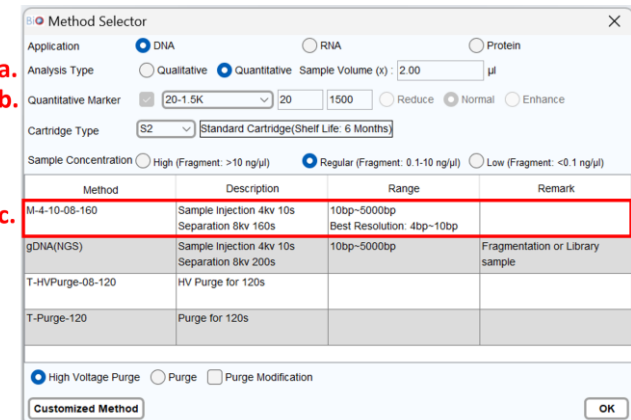
E. Software Operation Guide



1. Sample Position: Place the sample and select the corresponding position. Input sample information if necessary.



2. Method: Set (a) Analysis Type to Quantitative and input the sample volume for the sample mix. Select (b) Quantitative Marker and (c) Analytic Method in the Method Selector.



• Adjust injection conditions based on sample concentration.

Sample Concentration	High (2kV, 10s)	Regular (4kV, 10s)	Low (8kV, 10s)
Fragmented DNA	> 10 ng/μL	0.1-10 ng/μL	0.01-0.1 ng/μL
Smear DNA	> 50 ng/μL	2-50 ng/μL	0.5-2 ng/μL

3. Sample Duration: Adjust the sample injection time to increase/decrease injection amount.

• Modify injection conditions based on sample concentration.

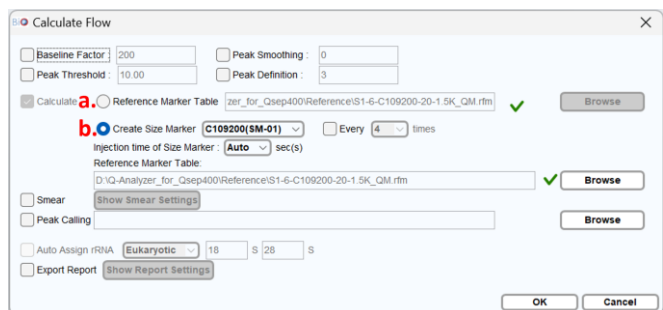
4. Runs: Set the repetition time.

5. Separation Duration: Adjust the duration to extend/reduce the separation time.

(Optional)

6. Result Name: Input the result name for the result file.

7. Para: Choose between (a) Reference Marker Table and (b) Create Size Marker for calculation.



• When using "Create Size Marker" function, select the appropriate size marker you use. For example, "20-1.5k" is paired with C109200, "20-5k" is paired with C109301.

8. Click "Run" to start the sequence analysis.

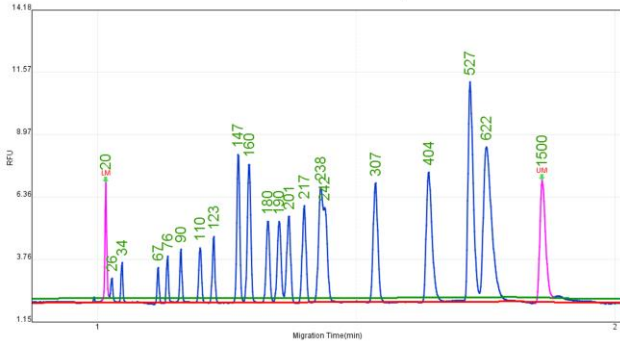
User Guide

4-Channel Standard Quantitative Cartridge Kit (C405101-Q)

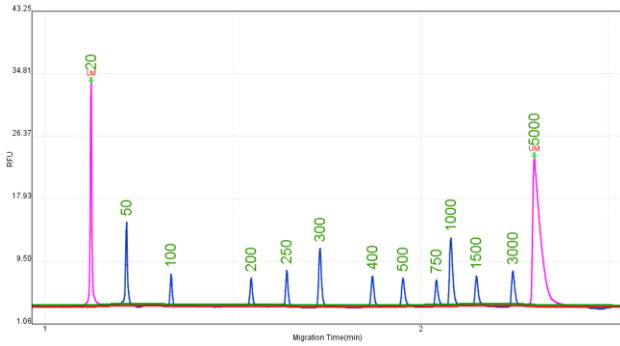
F. Result & Application

Quantitative Marker & Size Marker Result

20-1.5K & C109200 15-622 bp Size Marker



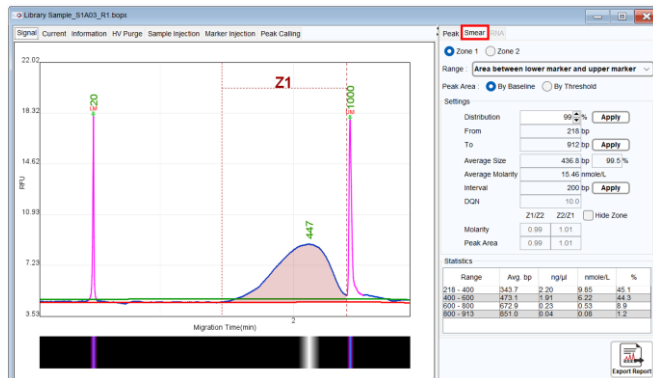
20-5K & C109301 50-3000 bp Size Marker



- The 4-channel system can be interpreted as 4 individual cartridges. Therefore, there should be 30-50% individual differences between each of them.

Fragmented & Library Sample Validation

Use the smear function to obtain average size, molarity, and size distribution of fragmented DNA or library DNA samples.



G. Troubleshooting

Before attempting any troubleshooting, ensure that the system is functioning properly and that all operations are following the instructions.

If encountering unstable current during sample injection or separation steps, which may be caused by unknown substances in PCR reagent buffer or other kit buffers, consider the following solutions:

- Dilute the sample using dilution buffer.
- Centrifuge the sample for a period to allow residues to accumulate at the bottom of the tube.
- Insert a "T-purge-120" method between several sample runs. For example, insert one run of "T-Purge-120" every 5-10 sample runs.

Sequence		Assay									
Open	Save	Save As									
SN	Sample Position	Method	Sample Duration	Runs	Separation Duration	Result Name		Para			
1	A-01.A...	M-4-10-08-160	10	1	160	Test 1		Sample ID			
2		T-Purge-120	0	1	0	Sample ID					
3	B-01.B...	M-4-10-08-160	10	1	160	Test 2		Sample ID			

H. Cartridge Disposal

Please wear gloves before discarding the cartridge.



- Bend the cartridge tip.
- Open the caps on the gel reservoirs and remove the inner caps.
- Pour the gel into the chemical waste container.
- Dispose of the cartridge in the trash bin.

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